

1 **High frequency of carbapenem-resistant *Acinetobacter baumannii***
2 **in patients with Diabetes Mellitus in Saudi Arabia**

3 Alsultan AA¹, Evans BA^{2*}, Elsayed EA¹, Al-Thawadi S³, Al-Taher AY¹, Amyes SGB⁴, Al-
4 Dughaym AM¹ and Hamouda A⁵.

5
6 ¹College of Medicine, King Faisal University, Alahsa 31982, P.O. Box 400, Kingdom of
7 Saudi Arabia; ²Department of Life Sciences, Faculty of Science and Technology, Anglia
8 Ruskin University, East Road, Cambridge, CB1 1PT, UK; ³King Faisal Specialist Hospital &
9 Research Centre, P. O. Box 3354, Riyadh 11211, Saudi Arabia; ⁴Medical Microbiology,
10 University of Edinburgh, The Chancellor's Building, 49 Little France Crescent, Edinburgh,
11 EH16 4SB, UK; ⁵Taibah University, Department of Clinical Biochemistry, Universities road,
12 Almadinah Almunawarah, P. O. Box 344, Saudi Arabia.

13

14 *Corresponding author. Email: benjamin.evans@anglia.ac.uk

15 Phone: +44 (0)845 271 3333

16

17 Contents Category: Clinical Microbiology and Virology

18 Running Title: *Acinetobacter* in diabetics in Saudi Arabia

19

20

21 **Abstract**

22 Carbapenem resistant *Acinetobacter baumannii* is becoming increasingly prevalent in
23 patients with diabetes mellitus in the Middle East. We examined the relationship between
24 these bacteria and their resistance mechanisms with the diabetic disease status of patients.
25 Susceptibilities of 271 isolates to carbapenems, tigecycline and colistin were determined,
26 followed by detection of carbapenemase genes. A *bla_{VIM}* gene was detected in ~95% isolates;
27 *bla_{OXA-23}* and *bla_{OXA-40}* genes were also prevalent. Diabetic patients were significantly more
28 likely to carry carbapenem-resistant isolates. Carbapenem resistant *A. baumannii* is a serious
29 problem in diabetics, and molecular detection of resistance mechanisms in these isolates is
30 required.

31

32

33

34

35

36

37

38

39

40

41

42

43

44

45

46 **Introduction**

47 One of the greatest threats to modern medicine is the increasing prevalence of antibiotic-
48 resistant bacteria, particularly Gram-negative bacteria (Boucher *et al.*, 2009). One of these
49 bacteria, *Acinetobacter baumannii*, has risen to prominence due to the international
50 dissemination of multidrug-resistant lineages resistant to the carbapenem antibiotics (Higgins
51 *et al.*, 2009; Towner *et al.*, 2008; Turton *et al.*, 2007). *A. baumannii* can become resistant to
52 the carbapenems through a number of mechanisms including expression of OXA-type
53 carbapenemases and expression of an acquired metallo- β -lactamase (Evans *et al.*, 2013;
54 Turton *et al.*, 2006). Recent studies have shown that the prevalence of carbapenem-resistant
55 *A. baumannii* can be incredibly high in the Middle East (Al Johani *et al.*, 2010; Mugnier *et*
56 *al.*, 2009).

57 Another growing health concern particularly prominent in the Middle East is the increase in
58 the number of people with diabetes mellitus (Danaei *et al.*, 2011). Diabetes mellitus has been
59 shown to be a significant risk factor in the acquisition of serious hospital-acquired infections
60 with *A. baumannii* (Metan *et al.*, 2009; Michalopoulos *et al.*, 2011; Prata-Rocha *et al.*, 2012).
61 The combination of increasing prevalence of diabetic patients and of carbapenem-resistant *A.*
62 *baumannii*, to which these patients appear to be particularly susceptible, presents a worrying
63 scenario of a rapidly rising number of difficult-to-treat infections. In order to begin to
64 understand the nature of this problem, we investigated the prevalence of carbapenem-resistant
65 *A. baumannii* and the β -lactamase genes they carry in both diabetic and non-diabetic patients
66 from hospitals across Saudi Arabia.

67

68 **Methods**

69 A total of 271 isolates preliminarily identified as *A. baumannii* obtained from patients from
70 intensive care units (ICUs) in hospitals in Saudi Arabia between 2008 and 2011 were selected

71 for inclusion in the study. Isolates were confirmed as *A. baumannii* using the Vitek compact
72 II system and detection of a *bla*_{OXA-51-like} gene by PCR (Woodford *et al.*, 2006). These 271
73 isolates comprised two distinct groups. One group of 196 isolates were selected for inclusion
74 due to initial identification as being carbapenem resistant, and of these, 84 were obtained
75 from patients with diabetes mellitus. Patients were defined as being diabetic if they were
76 insulin users and being treated for diabetes under the care of a hospital. The remaining 75 of
77 the 271 isolates were selected for inclusion at random without any prior knowledge about the
78 antimicrobial sensitivity of the isolates or the diabetic status of the patients they were
79 obtained from, with the exception that all isolates included in the study came from different
80 patients. Of the group of 75 randomly selected isolates, 20 were obtained from diabetic
81 patients. Detection of β -lactamase genes and insertion sequences was performed as described
82 previously (Ellington *et al.*, 2007; Poirel & Nordmann, 2006; Woodford *et al.*, 2006).
83 Antibiotic MICs were determined by the plate doubling dilution method according to British
84 Society for Antimicrobial Chemotherapy (BSAC) guidelines (Andrews, 2010). As no
85 breakpoint for tigecycline exists for *A. baumannii*, the breakpoint for Enterobacteriaceae was
86 used. For statistical analyses, isolates showing an intermediate level of antibiotic resistance
87 were grouped with resistant isolates. Analyses were performed in SPSS v.20 (SPSS Inc,
88 Chicago, IL, USA), and where appropriate were corrected for multiple testing (Benjamini &
89 Hochberg, 1995).

90

91 **Results and Discussion**

92 The isolates included in the study came from a total of 37 hospitals and a wide range of
93 samples including blood, respiratory, urine and wound. Of the 24 hospitals contributing more
94 than one isolate, 9 contributed isolates from both diabetic and non-diabetic patients. Analyses
95 were first performed on the data by dividing it into the isolates selected because they were

96 carbapenem resistant, and those selected randomly (Table 1). Combining the datasets for
97 analysis was not possible as the method of isolate selection would bias the results.
98 Determination of antibiotic MICs confirmed that carbapenem resistance was found in almost
99 all isolates selected for this trait, with only 6 of the 196 isolates (3%) having lost resistance to
100 both imipenem and meropenem during storage. Carbapenem resistance was also prevalent in
101 the randomly selected isolates, with a patient being more likely to carry a carbapenem-
102 resistant isolate than not (chi squared test, $\chi^2 = 7.053$, d.f. = 1, $p = 0.008$). A high proportion
103 of isolates (99%) carried genes for at least one acquired carbapenem-hydrolysing β -
104 lactamase, with *bla*_{VIM} sequences particularly prevalent. Of the 32 carbapenem-susceptible
105 isolates, 29 (91%) were positive for a *bla*_{VIM} gene.

106 The data were then analysed with respect to the disease status of the patient. Overall there is a
107 significantly higher proportion of diabetic patients in the carbapenem-resistant isolate group
108 than in the randomly selected isolate group (44% and 28% respectively, chi squared test, $\chi^2 =$
109 5.787, d.f. = 1, $p = 0.016$). The data were then sub-divided into those isolates that were
110 obtained from patients with diabetes, and those obtained from non-diabetics. Analyses were
111 performed on all categories for which there were enough data (Table 2). Amongst the
112 randomly selected isolates, diabetic patients were significantly more likely to carry an isolate
113 with a *bla*_{OXA-23} gene or for it to be carbapenem-resistant, consistent with the previous finding
114 above that diabetics are over-represented in the carbapenem-resistant isolate group. In the
115 carbapenem-resistant isolate group, diabetic patients were significantly more likely to carry
116 an isolate with an *ISAb*₂ or *ISAb*₃ insertion sequence, and significantly less likely to carry
117 a tigecycline-resistant isolate. A similar result was found in the randomly selected isolates. It
118 was also noted that there was a higher proportion of isolates carrying more than one gene of
119 *bla*_{OXA-23}, *bla*_{OXA-40} or *ISAb*₁ upstream of *bla*_{OXA-51} in non-diabetic patients than in those
120 with diabetes (10% and 1% respectively, chi squared test, $\chi^2 = 9.085$, d.f. = 1, $p = 0.003$).

121 In the present study we find that carbapenem resistance is very prevalent in *A. baumannii*
122 isolates from hospitals in Saudi Arabia, with isolates much more likely to be resistant than
123 not. Worryingly, a very high percentage of isolates carry a VIM-type metallo- β -lactamase
124 alongside high levels of carriage of the acquired carbapenemases *bla*_{OXA-23} and *bla*_{OXA-40}. As
125 has been noted previously for metallo- β -lactamases (Franklin *et al.*, 2006; Peleg *et al.*, 2005),
126 the number of isolates carrying a gene for a VIM-type enzyme is much higher than the
127 number of isolates phenotypically resistant to carbapenems, reinforcing the need for
128 molecular diagnostics particularly in regions where there is a known problem with such
129 organisms.

130 Another health issue that is particularly acute in the Middle East in countries such as Saudi
131 Arabia is the increasing number of patients diagnosed with diabetes mellitus (Danaei *et al.*,
132 2011). Previous studies have shown that diabetes is a significant risk factor in acquiring a
133 serious infection with *A. baumannii* (Metan *et al.*, 2009; Michalopoulos *et al.*, 2011; Prata-
134 Rocha *et al.*, 2012). To our knowledge, here we show for the first time that diabetic patients
135 with an *A. baumannii* infection are more likely than non-diabetic patients to carry a
136 carbapenem-resistant isolate. With the prevalence of diabetes increasing, this represents a real
137 healthcare problem as it amplifies the risk posed to diabetics from infection with *A.*
138 *baumannii*. Previously we demonstrated that particular *A. baumannii* clones were associated
139 with diabetic patients from Saudi Arabia (Alsultan *et al.*, 2009). In the present study there are
140 also indications that the isolates harboured by diabetics and non-diabetics differ. The further
141 characterisation of these isolates through typing and β -lactamase gene sequencing will allow
142 us to establish whether there are particular bacterial clones that are associated with diabetic
143 patients.

144

145

146 **Acknowledgements**

147 The authors would like to thank King Abdulaziz City for Science and Technology (KACST)
148 for funding, and to thank King Faisal University and the Vice President for Graduate Studies
149 and Scientific Research for their support. In addition, Hani Alrasasi, Hani Alfarhan, Hajer
150 Alduhilan, Nof Alhumaini, Fatemah Alnajar, Abdulrazaq Alwebari, Abdulkadir Abuli, and
151 Mohammed Alburake are thanked for their technical assistance. We would like to thank two
152 anonymous reviewers for their useful comments which helped clarify the manuscript.

153

154

155

156

157

158

159

160

161

162

163

164

165

166

167

168 **References**

169 **Al Johani, S. M., Akhter, J., Balkhy, H., El-Saed, A., Younan, M. & Memish, Z. (2010).** Prevalence of
170 antimicrobial resistance among gram-negative isolates in an adult intensive care unit at a
171 tertiary care center in Saudi Arabia. *Ann Saudi Med* **30**, 364-369.

172 **Alsultan, A. A., Hamouda, A., Evans, B. A. & Amyes, S. G. (2009).** *Acinetobacter baumannii*:
173 emergence of four strains with novel *bla*_{OXA-51-like} genes in patients with diabetes mellitus. *J*
174 *Chemother* **21**, 290-295.

175 **Andrews, J. M. (2010).** BSAC methods for antimicrobial susceptibility testing, version 9.1.

176 **Benjamini, Y. & Hochberg, Y. (1995).** Controlling the false discovery rate: a practical and powerful
177 approach to multiple testing. *J Roy Stat Soc B* **57**, 289-300.

178 **Boucher, H. W., Talbot, G. H., Bradley, J. S., Edwards, J. E., Gilbert, D., Rice, L. B., Scheld, M.,
179 Spellberg, B. & Bartlett, J. (2009).** Bad bugs, no drugs: no ESKAPE! An update from the
180 Infectious Diseases Society of America. *Clin Infect Dis* **48**, 1-12.

181 **Danaei, G., Finucane, M. M., Lu, Y., Singh, G. M., Cowan, M. J., Paciorek, C. J., Lin, J. K., Farzadfar,
182 F., Khang, Y. H. & other authors (2011).** National, regional, and global trends in fasting
183 plasma glucose and diabetes prevalence since 1980: systematic analysis of health
184 examination surveys and epidemiological studies with 370 country-years and 2.7 million
185 participants. *Lancet* **378**, 31-40. Epub 2011 Jun 2014.

186 **Ellington, M. J., Kistler, J., Livermore, D. M. & Woodford, N. (2007).** Multiplex PCR for rapid
187 detection of genes encoding acquired metallo-beta-lactamases. *J Antimicrob Chemother* **59**,
188 321-322.

189 **Evans, B. A., Hamouda, A. & Amyes, S. G. (2013).** The Rise of Carbapenem-Resistant *Acinetobacter*
190 *baumannii*. *Curr Pharm Des* **19**, 223-238.

191 **Franklin, C., Liolios, L. & Peleg, A. Y. (2006).** Phenotypic detection of carbapenem-susceptible
192 metallo-beta-lactamase-producing gram-negative bacilli in the clinical laboratory. *J Clin*
193 *Microbiol* **44**, 3139-3144.

194 **Higgins, P., Dammhayn, C., Hackel, M. & Seifert, H. (2009).** Global spread of carbapenem-resistant
195 *Acinetobacter baumannii*. *J Antimicrob Chemother.*

196 **Metan, G., Sariguzel, F. & Sumerkan, B. (2009).** Factors influencing survival in patients with multi-
197 drug-resistant *Acinetobacter* bacteraemia. *Eur J Intern Med* **20**, 540-544. Epub 2009 May
198 2029.

199 **Michalopoulos, A., Falagas, M. E., Karatza, D. C., Alexandropoulou, P., Papadakis, E., Gregorakos,**
200 **L., Chalevelakis, G. & Pappas, G. (2011).** Epidemiologic, clinical characteristics, and risk
201 factors for adverse outcome in multiresistant gram-negative primary bacteremia of critically
202 ill patients. *Am J Infect Control* **39**, 396-400. Epub 2010 Oct 2030.

203 **Mugnier, P. D., Bindayna, K. M., Poirel, L. & Nordmann, P. (2009).** Diversity of plasmid-mediated
204 carbapenem-hydrolysing oxacillinases among carbapenem-resistant *Acinetobacter*
205 *baumannii* isolates from Kingdom of Bahrain. *J Antimicrob Chemother* **63**, 1071-1073. Epub
206 2009 Feb 1027.

207 **Peleg, A. Y., Franklin, C., Bell, J. M. & Spelman, D. W. (2005).** Dissemination of the metallo-beta-
208 lactamase gene *bla*_{IMP-4} among gram-negative pathogens in a clinical setting in Australia.
209 *Clin Infect Dis* **41**, 1549-1556. Epub 2005 Oct 1531.

210 **Poirel, L. & Nordmann, P. (2006).** Genetic structures at the origin of acquisition and expression of
211 the carbapenem-hydrolyzing oxacillinase gene *bla*_{OXA-58} in *Acinetobacter baumannii*.
212 *Antimicrob Agents Chemother* **50**, 1442-1448.

213 **Prata-Rocha, M. L., Gontijo, P. P. & de Melo, G. B. (2012).** Factors influencing survival in patients
214 with multidrug-resistant *Acinetobacter baumannii* infection. *Braz J Infect Dis* **16**, 237-241.

215 **Towner, K. J., Levi, K. & Vlassiadi, M. (2008).** Genetic diversity of carbapenem-resistant isolates of
216 *Acinetobacter baumannii* in Europe. *Clin Microbiol Infect* **14**, 161-167.

217 **Turton, J. F., Gabriel, S. N., Valderrey, C., Kaufmann, M. E. & Pitt, T. L. (2007).** Use of sequence-
218 based typing and multiplex PCR to identify clonal lineages of outbreak strains of
219 *Acinetobacter baumannii*. *Clin Microbiol Infect* **13**, 807-815.

220 **Turton, J. F., Ward, M. E., Woodford, N., Kaufmann, M. E., Pike, R., Livermore, D. M. & Pitt, T. L.**
221 **(2006).** The role of IS_{Aba1} in expression of OXA carbapenemase genes in *Acinetobacter*
222 *baumannii*. *Fems Microbiol Lett* **258**, 72-77.

223 **Woodford, N., Ellington, M. J., Coelho, J. M., Turton, J. F., Ward, M. E., Brown, S., Amyes, S. G. B. &**
224 **Livermore, D. M. (2006).** Multiplex PCR for genes encoding prevalent OXA carbapenemases
225 in *Acinetobacter* spp. *Int J Antimicrob Ag* **27**, 351-353.

226

227

Characteristic	Carbapenem resistant		Random	
	No.	%	No.	%
Total isolates	196	100	75	100
No. diabetics	84	43	20	27
<i>bla</i> _{OXA-23}	108	55	42	56
<i>bla</i> _{OXA-40}	59	30	9	12
<i>bla</i> _{OXA-58}	0	0	0	0
ISAba1- <i>bla</i> _{OXA-51}	7	4	0	0
ISAba1	178	91	57	76
ISAba2	9	5	1	1
ISAba3	19	10	0	0
IS18	0	0	0	0
<i>bla</i> _{VIM}	182	93	72	96
Imipenem resistant	181	92	49	65
Meropenem resistant	186	95	49	65
Tigecycline resistant	25	13	9	12
Colistin resistant	2	1	0	0

228
229

230 **Table 1:** Presence of genotypic and phenotypic characteristics amongst isolates.

231

Carbapenem resistant	Diabetic		Non-diabetic		Chi Squared		
	No.	%	No.	%	statistic	d.f.	<i>p</i> value*
Total	84	43	112	57	-	-	-
<i>bla</i> _{OXA-23}	46	55	62	57	0.086	1	0.9228
<i>bla</i> _{OXA-40}	25	30	31	28	0.04	1	0.8410
<i>ISAba1</i>	78	93	97	89	0.839	1	0.5400
<i>ISAba2</i>	9	11	0	0	12.25	1	0.0028
<i>ISAba3</i>	16	19	3	3	14.193	1	0.0020
<i>bla</i> _{VIM}	79	94	100	92	0.374	1	0.7213
Imipenem resistant	80	95	98	90	1.88	1	0.2914
Meropenem resistant	83	99	100	92	4.822	1	0.0672
Tigecycline resistant	5	6	20	18	6.465	1	0.0440
Random							
Total	20	28	55	72	-	-	-
<i>bla</i> _{OXA-23}	16	76	26	47	5.14	1	0.0460
<i>bla</i> _{OXA-40}	0	0	9	16	3.719	1	0.0720
<i>ISAba1</i>	16	80	41	75	0.239	1	0.6250
Imipenem resistant	20	100	29	53	14.471	1	0.0006
Meropenem resistant	20	100	29	53	14.471	1	0.0006
Tigecycline resistant	0	0	9	16	3.898	1	0.0768

232
233 **Table 2:** Comparison of genotypic and phenotypic traits in diabetic and non-diabetic patients.
234 *Corrected *p* values according to Benjamini and Hochberg False Discovery Rate (Benjamini
235 & Hochberg, 1995), values in bold are significant at the 5% level.

236

237